

I'm not robot!

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Cap Talk e E Operations e -Aerospace Education f o year YEAGER TIPS OF PROMY I. Akers J. C., Gonda D., Kim R., Carter B. S. and Chen C. C. (2013). Biogan of Extracellular Vesicles (EV): Exosomes, microveses, retrovirus and apopthotic bodies. *J Neurooncol* 113 (1): 1-11. [PMC Free article] [PubMed] [Google Scholar] 2. Akou Adjei D., Johnson M. C. and Vogt V. M. (2005). The domain of the retroviral capsid determines the size of the Vion, the morphology and the assembly of the GAG in varchus -like particles. *J Virol* 79 (21): 13463-13472. [PMC Free article] [PubMed] [Google Scholar] 3. Balasubramaniam M. and Freed E. (2011). New ideas about the assembly and treated HIV. *FISIUIOLOGY* (Bethesda) August 26 (4): 236-251. [PMC Free article] [PubMed] [Google Scholar] 4. Block O., Mitra A., Novotny L. and Dykes C. (2012). A rude and free-free motion for quantification of human immunodeficiency particles type 1 by nanospectroscopy. *J Virol Methods* 182 (1-2): 70-75. [PMC Free article] [PubMed] [Google Scholar] 5. Eckwahl M.J., Telesnitsky A. and Wolin s.l. (2016). Host ma packaging by retrovirus: a newly sintended story. *MBio* 7 (1): E02025-02015. [PMC Free article] [PubMed] [Google Scholar] 6. Forster F., Medalia O., Zauberman N., Baumeister W. and Fass D. (2005). Structure of the protein complex of the envelope of rerovavirus in situ studied by cleaner tomography. *PROC NATL ACAD SCI U S A* 102 (13): 4729-4734. [PMC Free article] [PubMed] [Google Scholar] 7. Fujisawa R., Mcatee F. J., Favara C., Hayes S. F. and Portis J.L. (2001). The n-terminal cleavage fragment of the glycosylated gag is incorporated into the particles of Murino Oncornavirus. *J Virol* 75 (22): 11239-11243. [PMC Free article] [PubMed] [Google Scholar] 8. Fujisawa R., Mcatee F. J., Zirbel J.H. and Portis J.L. (1997). Characterization of the glycosylated gag expressed by a leukemia virus identification of differences in vitro and in vivo processing. *J Virol* 71 (7): 5355-5360. [PMC Free Article] PMC 9. Gardiner C., Shaw M., Hole P., Smith J., Tannetta D., Redman C. W. and Sargent I. L.(2014). Measurement of the refraction index by nanoparticles tracking analysis reveals heterogeneity in extracellular vesicles. *J Extracell Vesicles* 3: 25361. [PubMed free article] [Google Scholar]10. Houzet L., Gay B., Morichaud Z., Briant L. and Mougel M.(2006). Intracellular assembly and sprouting of the Murine Leukemia virus in infected cells. *Retrovirology* 3: 12. [PubMed free article] [Google Scholar]11. Kuji N., Yoshii T., Hamatani T., Hanabusa H., Yoshimura Y. and Kato S.(2008). Density and sedimentation dynamics of HIV-1 in two density-gradient media for semen processing. *Fertil Steril* 90(5): 1963-1967. [PubMed] [Google Scholar]12. Kurg R., Reinsalu O., Jagur S., Ounap K., Vosa L., Kasvandik S., Padari K., Gildemann K. and Ustav M.(2016). Biochemical and proteomic characterization of retrovirus-based microparticles Gag transporting melanoma antigens. *Sci Rep* 6: 29425. [PubMed free article] [Google Scholar]13. Lippe R.(2018). Flow virometry: a powerful tool to characterize virus functionally. *J Virol* 92(3). [PMC free article] [PubMed] [Google scholar]14. Longo P. A., Kavran J. M., Kim M. S. and Leahy D. J.(2013). Transfection of transient mammal cells with polyethylene(PFI). *Enzymol Methods* 529: 227-240. [PubMed free article] [Google Scholar]15. Madison M. N. and Okeoma C. M.(2015). Exosomes: Implications in HIV-1 Pathogenesis. *Virus* 7(7): 4093-4118. [PubMed free article] [Google Scholar]16. Martin J. L., Cao S., Maldonado J. O., Zhang W. and Mansky L. M.(2016). Different particle morphologies revealed through comparative parallel analysis of particles such as retrovirus. *J Virol* 90(18): 8074-8084. [PMC free article] [PubMed] [Google scholar]17. Mateescu B., Kowal E. J., van Balkom B. W., Bartel S., Bhattacharyya S.Buzas E. I., Buck A. H., de Candia P., Chow F. W., Das S., S., T. A., Fernandez-Messina L., Haderk F., Hill A. F., Jones J. C., Van Keuren-Jensen K. R., Lai C. P., Lasser C., Liegro I. D., Lunavat T. R., Lorenowicz M. J., Maas S. L., Mager I., Mittelbrunn M., Momma S., Mukherjee K., Nawaz M., Pegtel D. M., Pfaffl M. W., Schiffelers R. M., Tahara H., Thery C., Tosar J. P., Wauben M. H., Witwer K. W. and Nolte-t Hoen E. N.(2017). Obstacles and opportunities in the functional analysis of extracellular vesicle RNA- an ISEV position paper. *J Extracell Vesicles* 6(1): 1286095. [PMC free article] [PubMed] [Google Scholar]18. Melana S. M., Nepomnaschy L., Sakalian M., Abbott A., Hasa J., Holland J. F. and Pogo B. G.(2007). Characterization of viral particles isolated from primary cultures of human breast cancer cells. *Cancer Res* 67(18): 8960-8965. [PubMed] [Google Scholar]19. Molle D., Segura-Morales C., Camus G., Berliz-Torrent C., Kjems J., Basyk E., Bertrand E.(2009). Endosomal trafficking of HIV-1 gag and genomic RNAs regulates viral egress. *J Biol Chem* Jul 29: 19727-43 [PMC free article] [PubMed] [Google Scholar]20. Nestola P., Peixoto C., Silva R. R., Alves P. M., Mota J. P. and Carrondo M. J.(2015). Improved virus purification processes for vaccines and gene therapy. *Biotechnol Bioeng* 112(5): 843-857. [PubMed] [Google Scholar]21. E. Nolte-t Hoen, Cremer T., Gallo R. C. and Margolis L. B.(2016). Extracellular vesicles and viruses: Are they close relatives? *Proc Natl Acad Sci U S A* 113(33): 9155-9161. [PMC free article] [PubMed] [Google Scholar]22. O'Connor T. E., Rauscher F. J. and Zeigel R. F.(1964). Density gradient centrifugation of a murine leukemia virus. *Science* 144(3622): 1144-1147. [PubMed] [Google Scholar]23. Orenstein J. M., Meltzer M. S., Phipps T. and Gendelman H. E.(1988). Cytoplasmic assembly and accumulation of human immunodeficiency virus types 1 and 2 in recombinant human colony-stimulating factor-1-treated human monocytes: an ultrastructural study. *J Virol* 2578-2586. [PMC Free article] [PubMed] [Google Scholar] 24. Pang Y., Song H. and Cheng W. (2016). Using the typical trap to measure the refractions of an animal varchus in a high precise culture fluid. *Biomed Opt Express* 7 (5): 1672-1689. [PMC Free article] [PubMed] [Google Scholar] 25. Pillemer E. A., Koistra D. A., Witte O. N. and Weissman I. L. (1986). The monoclonal antibody is the amino-terminal sequence of gag polyprotections glycosilated glycosy of the leukemia of leukemia demonstrates its unusual orientation in the cell membrane. *J Virol* 57 (2): 413-421. [PMC Free article] [PubMed] [Google Scholar] 26. Quek C., Bellingham S. A., Jung C., Scicluna B. J., Shambok M. C., Sharples R. A., Cheng L. and Hill A. F. (2017). Defining the purity of the necessary exosomes for the small RNA diagnosis profile suitable for the discovery of biomarkers. *RNA BIOL* 14 (2): 245-258. [PMC Free article] [PubMed] [Google Scholar] 27. Raposo G., Moore M., Innes D., Leijendekker R., Leigh-Brown A., Benaroch P. and Geuze H. (2002). Human macrons accumulate HIV-1 particles in MHC II compartments. *TRAND* 3 (10): 718-729. [Pubmed] [Google Scholar] 28. Raposo G., Nijman H. W., Stoorvogel W., Liejendekker R., Harding C. V., Melief C. J. and Geuze H. J. (1996). B lymphons secrete vesicle presenters of Anthagenos. *J Exp Med* 183 (3): 1161-1172. [PMC Free article] [PubMed] [Google Scholar] 29. RayaproLu V., Ramsey J., Wang J.C. and Mukhopadhyay S. (2018). Purification of the Alfavirus using low speed centrifugation. *Bio-Protocol* 8 (6): E2772. [PMC Free article] [PubMed] [Google Scholar] 30. Renner T.M., Belanger K., Lam C., Gerpe M. C. R., McBane J. E. and Langlois M. A. (2018). Complete glycosylated gag of numb leukemia can be associated with viral envelope as a type I integral membrane protein 92 (6): E01530-17. [PMC Free article] [PubMed] [Google 31. Rosales Gerpe M. C., Renner T.M., Belanger K., Lam C., Aydin H. E LANCLOIS M. A. (2015). 52-11-78 sdohteM. tcnitsid yllanoitcnuf dna yllacimehoib was notartiifartlu laqifirtneq laitneuges yb 3668MIL enil lec recnac noloc namuh eht morf detaiosi selcisevorcim dehs dna semosoxe deifrup-02 L rentar dna _P namraepS, V ivakahtra [Speaking] [Sighs] J. niertop teN eht fo noisserpxe eht dna _gaG htw tcaretni ot yticacap sti _niertopocylg lariv eht fo noitazilacol ralullec eht yb denimreted si sepytdoesp lariworte fo ylbmess ecafirus lec susrev40. m. m. m., siljander p. r., andreu z., zavec a. b., borras f. e., buzas e. i., buzas k., couple e., cappello f., oak j., glues and., Lamb of the silva a. fais s., Falcon-Perez j. m. biological properties of extracellular vesicles and their physiological functions. *j extracell vesicles* 4: 27066. [pmc free article] [pubmed] [google Scholar]41. yeager m. Wilson-Kubalek e. m., weiner s. g., brown p. o. and reign A.(1998). suramolecular organization of immature and mature murina leukemia virus revealed by electronic cryomicroscopy: implications for retroviral assembly mechanisms. *proc natl acad sci u s a* 95(13): 7299-7304. [pmc free article] [pubmed] [google scholar]

